

## **Solutions for Cortical Cell Cultures**

(for E17 to P0 cultures)

### **HBSS (for dissection)**

(use within 1 month)

10X HBSS (Gibco #310-4180) 50 ml  
1 M Hepes (pH 7.4) 1.25 ml (=2.5mM)  
1 M Glucose 15 ml (=6.5 mg/ml=35 mM)  
100 mM CaCl<sub>2</sub> 5 ml (=1mM)  
100 mM MgSO<sub>4</sub> 5 ml (=1mM)  
1 M NaHCO<sub>3</sub> 2 ml (=4mM)

Add sterile dH<sub>2</sub>O to a total vol. of 500 ml.

### **Dissociation Medium (DM)**

(use within 1 month)

1 M Na<sub>2</sub>SO<sub>4</sub> 20.44 ml (or 40.88 mls of 0.5 M Na<sub>2</sub>SO<sub>4</sub>) (keep stock at room temp.)  
0.5 M K<sub>2</sub>SO<sub>4</sub> 15 ml (keep stock at room temp)  
1 M MgCl<sub>2</sub> 1.45 ml  
100 mM CaCl<sub>2</sub> 0.63 ml  
1 M Hepes (7.4) 250 µl  
1 M Glucose 5 ml  
Phenol Red (0.5%) 0.5 ml  
0.1N NaOH 0.5 ml

Add sterile dH<sub>2</sub>O to a total vol. of 250 ml.

**[Make fresh enzyme and inhibitor solutions immediately before dissection. For P0 cultures, add Ky/Mg to enzyme and inhibitor solutions]**

### **Enzyme soln.**

DM 10 ml  
Add cystein-HCl 3.2 mg  
Add Papain 200 units  
Mix and let dissolve for 15 min. (@ 37 C)  
pH w/ 0.1 N NaOH (approx. 6 drops)  
or O<sub>2</sub>/CO<sub>2</sub> (pink too basic/ yellow too acidic)  
Filter through 0.2 um syringe filter.

### **Heavy Inhib. soln.**

DM (37 C) 6 ml  
BSA 60 mg  
Trypsin Inhib. 60 mg  
Place at 37 C.  
Mix and pH w/ 0.1 N NaOH (approx 12 drops)  
or O<sub>2</sub>/CO<sub>2</sub> (pink too basic/ yellow too acidic)  
Filter through 0.2 um syringe filter.

**Light Inhib. soln.**

DM (37 C) 9 ml  
Heavy Inhib. 1 ml  
Filter through 0.2 µm filter.  
Place at 37 C.

**E17/P0 Serum-Free Media (Neurobasal+B27)**

Neurobasal (GIBCO without glutamine) 47.5 mls  
B27 supplement (GIBCO) 1 ml  
Glutamine (200 mM) 0.5 ml  
Pen/Strep (GIBCO#15140-122) 1 ml

---

**Notes:**

1. Use 3ml medium per 60 mm plate. Replace half the media every 6 days.
2. Ara-C may be added two days after plating to P0 cultures to prevent glial cell overgrowth.
3. Alternate media:

F12/DMEM media

100 mls

F12 47 mls  
DMEM (w/o glutamine) 47 mls  
PenStrep 1 ml  
Glutamine(200mM) 250µl  
Rat Serum 5 mls

L15 medium

L15 (w/o glutamine) 500 mls  
1M Glucose 15 mls  
1M NaHCO<sub>3</sub> 12.9 mls  
PenStrep 5 mls

4. Glucose concentrations typically used:

McConnell: 3g/liter (1:100 of 30% Glucose)  
Baraban: 5.5 g/liter  
Bading: 3.6 g/liter (20 mM Glucose; 1 M Glucose=186 g/liter)